INVESTIGATION OF THE PHYTOCHEMICAL COMPOSITION AND HEPATOPROTECTIVE EFFECT OF LEUCANTHEMOPSIS TRIFURCATUM GROWING IN LIBYA

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ABSTRACT

Leucanthemopsis trifurcatum is common to Mediterranean countries and widely used in traditional medicine especially for north African countries. Due to the scarcity of researches about the pharmacological activities of *L. trifurcatum*, we intended to screen the *L. trifurcatum* ethanolic extract (LEE) for hepatoprotective effects against paracetamol induced chronic toxicity. LEE is subjected to preliminary phytochemical and pharmacological screening. Hepatotoxicity is induced in rats by chronic oral administration of paracetamol (PCM) for 30 days. LEE (300 and 500 mg/kg), and silymarin (25 mg/kg) were administered orally, for 30 days, along with PCM to explore their hepatoprotective activities. Phytochemically, LEE revealed the presence of alkaloids, carbohydrates, and flavonoids as well as nine separated spots on TLC paper using toluene : ethyl acetate: formic acid (5:4:1) solvent system. The treatment with LEE (300 and 500 mg/kg, p.o.) caused hepatoprotective activity in a dose dependent manner through significant reduction of liver enzymes (AST, ALT, and ALP) compared to paracetamol toxicity. 500 mg/kg of LEE exerted a profound hepatoprotective action comparable to silymarine. This activity may be attributed to the synergistic action between the phytochemical constituents of LEE.

KEYWORDS: *Leucanthemopsis trifurcatum*, antinflammatory, hepatoprotective, analgesic, anticonvulsant, phytoconstituents.

INTRODUCTION

The *Leucanthemopsis* genus, golden flower in Greek, belongs to the Asteraceae family and includes about 300 species. This genus synthesizes and accumulates a variety of secondary metabolites such as pyrethroids, monoterpenoids, sesquiterpenoids, triterpenoids, flavonoids, coumarins, steroids, phenolics, purines, lipids, and aliphatic compounds⁽¹⁻⁵⁾.

Leucanthemopsis trifurcatum (Desf.) are annual or rarely biennial herbs with small yellow flowers and distributed in North African countries; Morocco, Algeria, Tunisia, and Libya^(6,7). The species, *Leucanthemopsis trifurcatum*, has been known in Algeria and Tunisia as "gueredfa" or "ouazouaza". The heads of plant flower are locally used as health diet such as "assida" or "chorba". This plant is used traditionally in Tunisia to treat constipation, hepatic disorders, intestinal transit problems and to ameliorate women pain after parturition. In Libyan traditional medicine, the aerial parts of *L. trifurcatum* are used as an antiinflammatory and antispasmodic agent^(1,8).

Many studies has demonstrated the potential medicinal effects including antibacterial, antifungal, anti-HSV1, and antioxidant effects for some Chrysanthemum species^(3,4,9-11), However, a few number of studies has investigated the biological activities and phytochemically analysed *L. trifurcatum* extracts^(1,3,8,5,12).

Correspondence and reprint request: Gamal A. Salem Department of Pharmacology, Faculty of Veterinary Medicine, Zagazig University, Zagazig-Egypt. Email: gamal_vet_85@yahoo.com To the best of our knowledge, the secondary metabolites of *L. trifurcatum* growing in Libya as well as its pharmacological activities have not been studied yet,. We carried out this study to analyze the phytoconstituents and investigate the hepatoprotective activity of Libyan *Leucanthemopsis trifurcatum*.

MATERIALS AND METHODS Collection and authentication of plant material:

For research and experiment, we collect fresh aerial parts of plant *Leucanthemopsis trifurcatum* from the basin of the Mediterranean Sea, Libya in May 2014, which was authenticated by Dr. Huda Elgubbi, Department of Botany, College of Science, Misurata University, Misurata, Libya.

Extraction:

The air-dried aerials parts of plant of *L. trifurcatum* were prepared as a coarse powder and 500 g of this powdered material was macerated with distilled water and 95% w/v alcohol separately for 24 hrs and 72 hrs, respectively. Then, the extract were filtered through a muslin cloth and the resultant filtrates were concentrated under reduced pressure and vacuum dried. The yield of ethanolic extract was 13 % w/w. The extract were reconstituted in their extraction solvent to give the required concentration needed in this study. Freeze-dried extracts were collected in small glass bottles and kept at 30° C for further evaluation.

Preliminary Phytochemical Screening and thin Layer Chromatography (TLC):

Preliminary phytochemical screening for the detection of various phytoconstitunts such as alkaloids, carbohydrates, steroids, flavonoids, saponins, and glycosids was carried out by using standard procedures described by Harborne *et al*⁽¹³⁾. Thin layer chromatography studies of the extracts of *L. trifurcatum* carried out in various solvents at 30^o C using pre-coated plates silica gel 60 F₂₅₄, 7X6 cm (Merck)as adsorbent and the R_fvalues were determined.

Experimental Animals:

Thirty male and female Wistar rats (90-110 days) weighing 200-300 g. The animals were divided into groups and kept in plastic cages (47 * 34 * 18 cm) under a 12 h light/12 h dark cycle at room temperature (22 °C), with free access to standard ration and water. Animal care and the experimental protocol followed the principles and guidelines suggested by Faculty of Pharmacy-Misurata University and were approved by the local ethical committee.

Hepatoprotective Activity:

For evaluation the hepatoprotective activity of LEE against paracetamol-induced liver injury in experimental rats, 30 rats were used and randomly assigned to 5 groups (6 animals per group) and treated orally for 30 consecutive days. Group 1 served as normal control and received tween 80 (1%) (10 ml/kg body weight); groups 2 served as hepatotoxic control and received Paracetamol (PCM) (500 mg/kg); group 3 served as standard drug treatment group and received silymarin (25 mg/kg) + 500 mg/kg PCM; groups 4 received LEE (300 mg/kg) + 500 mg/kg PCM; groups 5 received LEE (500 mg/kg) + 500 mg/kg PCM. PCM was administered one hour before the administration of either silvmarin or the LEE At the end of the treatment period all animals were eusthetized with ether and blood was collected into plain dry tubes and centrifuged at 2,000 rpm for 10 min.

Biochemical analysis:

The serum samples obtained were transferred into Eppendorff tubes and were analyzed by INTEGRA 400 plus for estimation of liver enzymes: aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline Phosphatase (ALP), using commercially available kits (Roche Diagnostics GmbH, Mannheim, Germany).

Statistical Analysis:

The results obtained were presented as mean \pm standard error (SE). The significance of differences between means were analyzed statistically with one way analysis of variance (ANOVA; 95% confidence interval) and LSD post hoc tests using PSPP program (Linux operating system). Values of p<0.05 were taken to imply statistical significance.

RESULTS

Preliminary Phytochemical Screening and thin Layer Chromatography (TLC):

The preliminary phytochemical screening for the *L. trifurcatum* extracts showed the presence of Alkaloids, carbohydrates, and flavonoids. The tests also revealed that the absence of glycosids, saponins, steroids in LEE (table 1).

Constituents	CEE
Alkaloids	+ve
Carbohydrates	+ve
Glycosids	-ve
Flavonoids	+ve
Saponins	-ve
Steroids & Terpenes	-ve

Keys: +ve (Present), -ve (Absent).

Thin layer chromatography of the extracts of *L. tri-furcatum* was carried out using toluene: ethyl acetate: formic acid (5:4:1) as mobile phase respectively and the *Rf* values were recorded (table 2). The peaks were visualised at a wavelength of 366 nm in UV chamber.

(Table 2) Rf values of TLC solvent system for CEE.

Solvent system	No. of spots	$\mathbf{R}_{\mathbf{f}}$ values	Visualizing agent
Toluene: ethyl acetate: formic acid (5:4:1)	09	0.13, 0.17, 0.48, 0.55, 0.615, 0.73, 0.82, 0.88, 0.92.	UV-366 nm

Hepatoprotective Effect:

Chronic oral PCM (500 mg/kg) induced significant elevation in hepatic serum markers, AST, ALT, and ALP in rats. Meanwhile, oral administration of LEE (300 and 500 mg/kg) together with PCM reduced significantly, in a dose dependant manner, the increase in these hepatic markers in comparison to PCM treated group. Treatment with 500 mg/kg of LEE showed non significant differences with SLM (25 mg/kg) in serum AST, ALT, and ALP levels. These results are shown in (table 3).

(**Table 3**) Effect of LEE (300 and 500 mg/kg), and Silymarin (SLM) (25 mg/kg) on serum hepatic markers in PCM-induced chronic hepatotoxicity in rats:

	Liver Function Test		
Group	ALT (U/L)	ALP (U/L)	AST (U/L)
Control	$\begin{array}{c} 51.20 \pm \\ 6.76^a \end{array}$	$\begin{array}{c} 82.00 \pm \\ 16.81^a \end{array}$	85.00 ± 11.18^{a}
LEE 300 mg/kg + PCM (500 mg/kg)	$\begin{array}{c} 93.00 \pm \\ 4.58^b \end{array}$	$\begin{array}{c} 219.80 \pm \\ 9.36^b \end{array}$	652.60 ± 175.11 ^b
LEE 500 mg/kg + PCM (500 mg/kg)	76.80 ± 5.40°	${\begin{array}{c} 150.00 \pm \\ 20.26^{c} \end{array}}$	510.20 ± 60.57°
SLM (25 mg/kg) + PCM (500 mg/kg)	65.40 ± 9.76°	120.60 ± 36.15 ^c	415.40 ± 49.22°
PCM (500 mg/kg)	155.60 ± 16.33^{b}	229.40 ± 24.29^{d}	$\begin{array}{c} 933.60 \pm \\ 191.87^{d} \end{array}$

Different letters indicate significant difference between means result ($p \ge 0.05$).

DISCUSSION

Natural products and their derivatives have been known for many years as a source of therapeutic agents and of structural diversity⁽¹⁴⁾. A bout 40% of all medicines is either natural products or their sem-

isynthetic derivatives. Clinical, pharmacological, and chemical studies of these traditional medicines, which were derived predominantly from plants, were the basis of most early medicines such as aspirin, digitoxin, morphine, quinine, and pilocarpine⁽¹⁵⁾. The phytochemical screening of L. trifurcatum has shown the presence of various bioactive constituents^(2,3) which are responsible for most pharmacological activities of plants⁽¹⁶⁾. The results of the present study also supplement the folkloric usage of the L. trifurcatum which contain numerous known and unknown bioactive compounds with bio-activity. Thin layer chromatography (TLC) is particularly valuable for the preliminary separation and determination of plant constituents. The chromatographic profile may serve as a characteristic finger print for qualitative evaluation of L. trifurcatum.

In the present study, the ethanol extracts of aerial parts of L. trifurcatum was observed to exhibit hepatoprotective effect as demonstrated by a significant decrease in serum marker liver enzymes AST, ALT, and ALP level in rat induced with paracetamol hepatotoxicity. Analgesic drug paracetamol is commonly used model for screening hepatoprotective drugs. It has been shown that high doses of paracetamol produce centrilobular liver necrosis in in experimental animals and human^(17,18). For screening of hepatoprotective agents, paracetamol-induced hepatotoxicity has been used as a reliable method. Paracetamol is primarily metabolized by sulfation and glucuronidation, but as the dose is increased, these pathways become saturated and a greater proportion of the drug is available for oxidation by the microsomal cytochrome P-450 system. Moreover, induced liver toxicity and cell death is due to formation of toxic metabolites when a part of paracetamol is activated by hepatic cytochrome P-450 to a highly reactive metaboliteN-acetyl-p-benzoquinoneimine.

(NAPQI)^(19,20). NAPQI produces oxidative stress and cause glycogen and glutathione depletion by irreversible conjugation with sulfhydral groups of glutathione, which leads to increased lipid peroxidation by abstracting hydrogen from a polyunsaturated fatty acid and ultimately, liver damage due to higher doses of paracetamol^(21,22) The glutathione protects hepatocytes by combining with the reactive metabolite of paracetamol, thus preventing covalent binding to liver proteins⁽³³⁾. Reactive metabolites can exert initial cell stress through a wide range of mechanisms including depletion of glutathione (GSH) or binding to enzymes, lipids, nucleic acids and other cell structures⁽²⁴⁻²⁶⁾.

In the estimation of liver damage by paracetamol, the determination of these enzyme levels, especially ALT is largely used. It is generally acknowledged that administration of paracetamol caused a significant elevation of enzymes level such as AST, ALT and ALP, it has been attributed to damage structural integrity of liver, because they are cytoplasmic in location and released into circulation after cellular dam-

ages indicating development of hepatotoxicity. [27-39]. However, administration of the crude ethanol extract of *L. trifurcatum* whole plant (LEE) at various 300 and 500 mg/kg doses mediated a reduction in the levels of AST, ALT and ALP towards the normal value. This indicates a stabilization of plasma membrane as well as repair of hepatic tissue damage caused by paracetamol. This effect is in consonance with the common view that serum level of transaminases returns to normal following healing of hepatic parenchyma and regeneration of hepatocytes⁽³⁰⁾. Silymarin is a well-known hepatoprotective com-

pound isolated from Silybum marianum is reported to possess a protective effect on plasma membrane of hepatocytes and have multiple inhibitory multiple mechanisms of actions against different hepatotoxic agents. The antioxidant property and cell regenerating functions as effect of increased protein synthesis were considered as most important actions of silymarin⁽³¹⁾. Our study shows that ethanolic extracts of aerial parts of L. trifurcatum at higher dose (500 mg/kg) is comparable with standard drug Silymarin. Hepatoproective activites of LEE may be due to the antioxidant potential of flavonoid compounds as has been demonstrated from Chrysanthemum trifurcatum⁽¹⁾ and some other closely related plant species such as Chrysanthemum balsamita^(32,33), Chrysanthemum fontanesii⁽³⁴⁾, Chrysanthemum indicum^(34,35). Moreover, a number of scientific studies of other plants indicated that contain flavonoids, triterpenoids and steroids have protective effects on liver due to its antioxidant properties(36-42).

REFERENCES

1- Mokaddem-Daroui H, Touafek O, Kabouche A, Kabouche Z, Calliste C A, and Duroux J L. The Components and Antioxidant Activity of the Polar Extracts of Chrysanthemum Trifurcatum. Chemistry of Natural Compounds 2012; 48(3):498-499.

2- Kumar A, Singh S P, and Bhakuni R S. Secondary metabolites of Chrysanthemum genus and their biological activities. Curr. Sci.2005; 89(9): 1489-1501.

3- Ahlem B S, Fethia H S, Nathalie B, and Mahjoub A. Antimicrobial activities of four Tunisian Chrysanthe mum species. Indian J. Med. Res.2008; 127:183-92.

4- Lograda Takia, Ramdani Messaoud, Chalard Pierre, Figueredo Gilles, and Silini Hafsa. Chemical composition, antibacterial activity and chromosome number of Algerian populations of two Chrysanthemum species. Journal of Applied Pharmaceutical Science, suppl. Supplement: 2013; 13(8): S6-S11.

5- Sassi AB, Skhiri FH, Chraief I, Bourgougnon N, Hammami M, and Aouni M. Essential oils and crude extracts from *Chrysanthemum trifurcatum* leaves, stems and roots: chemical composition and antibacterial activity. Journal of Oleo Science [J Oleo Sci] 2014; Vol. 63 (6), pp. 607-17.

6- The Bulletin of The Natural History Museum 1993 Botany Series, Vol. 23, No. 2, pp. 55-177.

7- Haouas D, Ben Halima-Kamel M, Harzallh-Skhiri F, and Ben Hamouda M H. Assessment of insecticidal effect of *Chrysanthemum sp.* essential oils against *Tri*-

bolium confusum du Val (Coleopter, Tenebrionidae). The African of Plant Science and Biotechnology 2013

8- [Effects of the methanolic extract of Chrysanthemum trifurcatum (Desf.) Batt. and Trab. on rat duodenal motility]. C. R. Biol.C R Biol 2007 Mar 8;330(3):226-30. Epub 2007 Dec 8. Ahlem Ben Sassi, Fethia Harzallah-Skhiri, Wahida Borgi, Nabil Chouchène, Mahjoub Aouni

9- Andogan, B. C.; Baydar, H.; Kaya, S.; Demirci, M.; Oz- basar, D.; Mumcu, E. Antimicrobial activity and chemi- cal composition of some essential oils. Arch. Pharm. Res. 25, 860-864 (2002).

10- Khallouki F, Hmamouchi M, Younos C, Soulimani R, Bessiere J M, Essassi E M. Antibacterial and molluscicidal activities of the essential oil of Chrysan- themum viscidehirtum Fitoterapia 2000; 71: 544-546. (abstract)

11- Shafaghat A, Larijani K, Salimi F. Composition and antibacterial activity of the essential oil of Chrysanthemum parthenium flower from Iran. J. Essent. Oil.2010; Bear Pl. 12 (6) :708-713.

12- Ahlem Ben Sassi, Fethia Harzallah-Skhiri, Imed Chraief, Nathalie Bourgougnon, Mohamed Hammami, Mahjoub Aouni. Chemical composition and antimicrobial activities of the essential oil of (Tunisian) *Chrysanthemum trifurcatum* (Desf.) Batt. and Trab. Flower heads. C. R. Chimie 11 (2008) 324-330.

13- J.B. (Jeffrey B.. Harborne, Phytochemical methods : a guide to modern techniques of plant analysis 3rd ed, Chapman and Hall, 1998.

14- Kokate CK, A text book of Practical Pharmacognosy, Vallabh Prakashan 5th edition 2005 New Delhi India, 1994, 107–111

15- Peach and Tracey MV. In Modern Methods of plants analysis. Spingler and Vena Publishers Berlin 1935.

16- The Success of Natural Products in Drug Discovery: Mouhssen Lahlou, Pharmacology & Pharmacy, 2013, 4, 17-31.

17- Vermeulen NPE, Bessems JGM, Vande Streat R. Molecular aspects of paracetamol hepatotoxicity and it mechanism based prevention. Drug Metab Rev 1992; 24: 367-407.

18- Moore M, Thor H, Moore G, Nelson S, Moldeus P, Correnius S: The toxicity of acetaminophen and N-acetyl P-benzoquinone imine in isolated hepatocytes is associated with thio depletion and increased cytosolic Ca2+. J Biol Chem 1985, 260:13035–13040.

19- Wong LT, Whitehouse LW, Solemonraj G, Paul CJ: Pathways of Acetaminophen conjugate in the mouse. Toxicity Lett 1981, 9:145–151.

20- Savides MC, Oehne FW: Acetaminophen and its toxicity. J App Toxicol 1983, 3:95–111.

21- Savides M.C. and Oehme F W J, (Acetaminophen and its toxicity Appl. Toxicol, 1983;3 (2):95-111.

22- Mitchell J R, Jollow D J, Potter W Z, Gillette J R and Brodie, B B J, (Acetaminophen-Induced Hepatic Necrosis. Iv. Protective Role Of Glutathione), Pharmacol Exp Ther,1973; 187 (1): 211-217.

23- Tirmenstein MA, Nelson SP: Sub cellular binding and effects on calcium homeostasis produced by acetaminophen and a non-hepatotoxic region isomer 3hydroxyacetanilide in mouse liver. J Biol Chem 1989, 264:9814–9819. 24- Grypioti AD. Liver oxidant stress induced by paracetamol overdose. Internet J Pharmacol. 2006;4(2):p7.

25- Kozer E, Evans S, Barr J, Greenberg R, Soriano I, Bulkowstein, M Petrov, I. Chen-Levi, Z., Barzilay, B. and Berkovitch, M. Glutathione, glutathionedependent enzymes and antioxidant status in erythrocytes from children treated with high-dose paracetamol: Br J Clin Pharmacol.2003; 55(3): 234-40.

26- Pauli-Magnus C, Stieger B, Meier Y. Kullak-Ublick G A and Meier P J. (Enterohepatic transport of bile salts and genetics of cholestasis) J. Hepatol, 2005; 43(2): 342-357.

27- Hepatoprotective activity of Spilanthes paniculata flower extracts on liver damage induced by paracetamol in rats Syed A. Ayaz*, Shukla Mahanand, Subur W. Khan, Der Pharmacia Sinica, 2012, 3 (6):738-744].

28- Girish C, Koner BC, Jayanthi S, Rao KR, Rajesh B and Pradhan SC. Hepatoprotective activity of six polyherbal formulations in paracetamol induced liver toxicity in mice, Indian J Med Res 2009, 129(5): 569- 578.

29- Gutiérrez R M P and Solís R V. Rec. (Hepatoprotective and Inhibition of Oxidative Stress in Liver of *Prostechea michuacana*) Nat. Prod, 2009; 3(1): 46-51.

30- Thabrew MI, Joice PD, Rajatissa WA: Comparative study of the efficacy of *Paetta Indica* and *Osbeckia octandra* in the treatment of liver dysfunction. Planta med 1987; 53: 239 – 241.

31- Jalil Ur Rehman, Najam Us Saqib, Naveed Akhtar, Muhammad Jamshaid, Hafiz Muhammad Asif, Sabira Sultana and Riaz Ur Rehman: Hepatoprotective activity of aqueous-methanolic extract of Suaeda fruticosa in paracetamol-induced hepatotoxicity in rabbits. Bangladesh J Pharmacol 2013; 8: 378-381

32- Aneta T. Popova 1 Dasha S. Mihaylova 2 Iordanka N. Alexieva: The effect of freezing on the antioxidant activity of Bulgarian *Chrysanthemum balsamita*. J. BioSci. Biotech. 2014, SE/ONLINE: 17-21.

33- Daniela Benedec1, Lorena Filip1, Laurian Vlase1, Constantin Bele, Bogdan Sevastre, Oana Raita, Neli-Kinga Olah and Daniela Hanganu: *In vitro* study of antioxidant activity and phenolic content of *Chrysanthemum balsamita* varieties. Pak. J. Pharm. Sci., Vol.29, No.4(Suppl), July 2016, pp.1359-1364.

34- Amel Amrani, Nassima Boubekri, Ouahiba Benaissa, Djamila Zama, Fadila Benayache, Samir Benayache. Ethanol Induced Toxicity and Lipid Peroxidation in Pregnant Mice: Protective Effects of Butanolic Extract from Leaves of Chrysanthemum fontanesii, Vitamin E and C. Journal of Stress Physiology & Biochemistry, Vol. 10 No. 2 2014, pp. 35-43 ISSN 1997-0838.

35- Trishna Debnath, Hai Lan Jin, Md Abul Hasnat, Yunsuk Kim, Nadira Binte Samad, Pyo-Jam Park And Beong Ou Lim. Antioxidant Potential And Oxidative Dna Damage Preventive Activity Of *Chrysanthemum Indicum* Extractsj.Journal Of Food Biochemistry Issn 1745-4514 440.

36- Leqin Ke and Haiyan Chen: Enzymatic-Assisted Microwave Extraction of Total Flavonoids from Bud of Chrysanthemum indicum L. and Evaluation of Biological Activities. Int. J. Food Eng. 2016; 12(6): 607–613.

37- Simon Rama Parmar, Patel Hitesh Vashrambhai, Kiran Kalia: Hepatoprotective Activity Of Some Plants Extract Against Paracetamol Induced Hepatotoxicity In Rats. Journal Of Herbal Medicine And Toxicology 4 (2) 101-106 (2010).

38- Syed A. Ayaz*, Shukla Mahanand, Subur W. Khan. Hepatoprotective activity of *Spilanthes paniculata* flower extracts on liver damage induced by paracetamol in rats. Der Pharmacia Sinica, 2012, 3 (6):738-744.

39- Mohsin Ali, M. Imran Qadir, Mohammad Saleem, Khalid Hussain Janbaz, Humaira Gul1, Liaqat Hussain and Bashir Ahmad. Hepatoprotective potential of Convolvulus arvensis against paracetamol-induced hepatotoxicity. Bangladesh J Pharmacol 2013; 8: 000-000.

40- Mohamed Sakran , Yasser Selim , and Nahla Zidan. A New Isoflavonoid from Seeds of *Lepidium sativum* L. and Its Protective Effect on Hepatotoxicity Induced by Paracetamol in Male Rats. *Molecules* 2014, *19*, 15440-15451. 41- Jalil Ur Rehman, Naveed Akhtar, Muhammad Younus Khan, Khalil Ahmad, Mukhtiar Ahmad, Sabira Sultana, Hafiz Muhammad Asif. Phytochemical Screening and Hepatoprotective Effect of *Alhagi maurorum* Boiss (Leguminosae) Against Paracetamol-Induced Hepatotoxicity in Rabbits. Tropical Journal of Pharmaceutical Research June 2015; 14 (6): 1029-1034.

42- Amina El-Shaibany, Molham AL-Habori, Shaza Al-Massarani, Ali El- Gamal, Ali Al-Ajami5 and Adenan Al-Adhl. Hepatoprotective Effect of *Pandanus odoratissimus* L Inflorescence Extracts in Acetaminophentreated Guinea Pigs. Tropical Journal of Pharmaceutical Research February 2016; 15 (2): 259-266.